

Report of the ISHLT Working Group on primary lung graft dysfunction Part IV: Prevention and treatment: A 2016 Consensus Group statement of the International Society for Heart and Lung Transplantation



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Dysfunction of the pulmonary allograft with impaired oxygenation and radiographic opacities consistent with pulmonary edema occurs in up to 50% of lung transplant (LTx) recipients.¹ In 2005, 4 severity grades of primary graft dysfunction (PGD 0, 1, 2 and 3) were defined by a working group within the International Society for Heart and Lung Transplantation (ISHLT).¹ In most cases, the injury is mild and transient, but in 25% to 30% of cases it can result in severe hypoxemia with a partial pressure of oxygen/fraction of inspired oxygen ratio (PaO₂/FIO₂) of <200 mm Hg (PGD 3) within the first 72 hours (T0 to T72) after LTx.^{2,3}

Despite advances in our understanding of donor and recipient risk factors, donor-recipient matching, organ preservation, surgical techniques and peri-operative care, PGD still accounts for significant morbidity and mortality after LTx. This Consensus Statement on prevention and treatment of PGD aims to update the previous publication from 2005⁴ by reviewing published evidence on novel strategies for reducing the incidence of PGD and for attenuating its severity once developed, in order to mitigate both short- and long-term PGD-related morbidity and mortality.

Prevention of PGD

Potential strategies to prevent and minimize the development and severity of PGD include: (1) optimizing donor

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and recipient selection, donor–recipient matching and management of donors and recipients pre-operatively; (2) improving lung preservation and storage techniques; and (3) improving lung implantation and reperfusion techniques, all based on currently known PGD risk factors.

Donors and recipients

Donor selection

Lungs are fragile and more sensitive to trauma compared with other organs. They may be acutely injured in the hours before and after brain death. This results from direct trauma due to: contusion; the resuscitation maneuver utilized; neurogenic edema; pulmonary emboli with thrombi or fat; aspiration of blood or gastric contents with infection; or ventilator-associated trauma and pneumonia. Any or all of these insults make lungs less suitable for transplantation compared with kidney and liver.^{5,6} As a result, only 10% to 15% of cadaveric multi-organ donors provide lungs believed to be suitable for transplantation according to criteria that were defined during the early days of successful LTx.⁷

Efforts to expand the donor pool by transplanting extended-criteria donor (ECD) lungs have occurred worldwide.⁸ Most reports demonstrate equivalent outcomes after ECD LTx with regard to PGD incidence, early and late outcomes and freedom from bronchiolitis obliterans syndrome (BOS). Three studies,^{8–10} however, have reported a higher incidence of PGD 3, with increased mortality in 2 studies.^{9,10} On the other hand, excluding these donors based solely on inherent donor risk factors, such as older age or considerable smoking history, may increase waitlist mortality, while the outcome for the great majority of recipients will still be much better than having not received a transplant.^{11–13} On balance, caution and clinical judgment are warranted when using ECD donor lungs with more than 1 extended criterion.¹⁰

Lungs recovered from controlled donors after circulatory death (DCD) are increasingly being transplanted, mainly in Australia, Canada and some European countries (including Belgium, The Netherlands, Spain, Switzerland, and the UK). In several institutional reports, early and late survival rates were comparable after DCD LTx and conventional donors after brain death (DBD) LTx. This was confirmed recently both in an analysis from the DCD Registry¹⁴ of the ISHLT and in a meta-analysis of 11 reported observational cohort studies.¹⁵ Five studies reported on PGD data for DCD and DBD cohorts. None of these reported significant differences in PGD rates between DCD and DBD cohorts. Also, in a pooled analysis of “controlled” (Maastricht Class III or IV) DCD LTx, there was no difference in PGD rates between DCD and DBD recipients.¹⁵ However, in the Madrid series, the incidence of PGD 3 in lung recipients from “uncontrolled” (Maastricht Class I or II) DCDs was reported to be as high as 38%, with serious impact on early- and mid-term mortality.¹⁶ Therefore, pre-transplant evaluation of Maastricht Class I or II donor lungs with ex-vivo

lung perfusion (EVLV) before LTx is now recommended by the Madrid group.¹⁷

Current evaluation of donor lung quality with gas exchange, chest radiograph and bronchoscopy is often difficult and quite subjective. Whenever possible, lungs should be inspected at the donor hospital. Gas exchange should be re-evaluated with the chest open and lungs fully ventilated. It is hoped that, in the near future, biomarkers correlating with (non-apparent) donor lung injury as well as reassessment with EVLP will help to better differentiate pulmonary allografts that should be declined or first treated, either in the donor before retrieval or during ex-vivo reconditioning.

Donor management

Donor management should be considered as a continuation of critical care after confirmation of brain death, with a shift in focus toward optimal continued functioning of individual organs.¹⁸ Comprehensive donor management by qualified personnel based on protocols will increase the quantity and quality of transplantable organs.^{19,20}

A systematic approach to respiratory management should be followed—in addition to hemodynamic management, hormonal resuscitation, electrolytes and fluid control and body temperature maintenance—in order to maximize the number of suitable donor lungs. In the past, respiratory management protocols used a non-protective ventilation strategy based on a tidal volume (TV) of 10 to 15 ml/kg body weight.²¹ More recent approaches to optimize lung recovery include: alveolar recruitment using high levels of positive end-expiratory pressure (PEEP) (15 cmH₂O); inspiratory pressures of 25 cmH₂O; bronchoscopy to assess and minimize respiratory secretions; 30° head elevation; and endotracheal cuff pressures of 25 cmH₂O to limit aspiration.^{22,23} After reporting that ventilation with higher TV was an independent risk factor for the development of acute lung injury,²⁴ Mascia et al were the first to prospectively study the impact of a new protective ventilation strategy on the number of lung donors.²⁵ This ventilation strategy is characterized by smaller TV (6 to 8 ml/kg predicted body weight) and lower PEEP (8 to 10 cmH₂O), continuous positive airway pressure during the apnea test and recruitment maneuvers, and is in accordance with the current standard of care for patients with acute respiratory distress syndrome (ARDS).²⁶ These different strategies were implemented in a general protocol that included: low TV (8 ml/kg); low PEEP (8 to 10 cmH₂O); recruitment maneuvers (PEEP of 15 to 18 cm H₂O); fluid restriction (targeted central venous pressure 6 to 8 mm Hg and extravascular lung water <10 ml/kg); use of diuretic, if necessary; addition of methylprednisone (15 mg/kg body weight); and thyroid supplementation in patients on inotropes. This protocol improved lung utilization without jeopardizing the acceptance rate for kidney grafts.²⁷ In a further study by Minambres et al, comparing 2 study periods before and after implementing an intensive lung donor management protocol, the rate of lung DBDs increased

significantly from 20.1% to 50%, thus quadrupling the total number of pulmonary grafts retrieved and patients receiving a lung transplant.²⁸ Of note, no differences were observed in early recipient survival or in the rate of PGD 3.

In a placebo-controlled, randomized trial, thyroid hormone alone or in combination with corticosteroids had no effect on donor lung function or yield, whereas steroids reduced progressive lung water accumulation.²⁹ The BOLD study, a randomized, placebo-controlled trial, evaluated the effect of nebulized albuterol on pulmonary edema, but failed to demonstrate any differences in donor oxygenation or lung utilization.³⁰

Recipient selection

Recipient selection criteria for LTx were recently revisited by an ISHLT Pulmonary Council working group.³¹ Since the previous publication in 2005,³² additional studies examining recipient-related risk factors have consistently reported an association of PGD with pre-transplant diagnosis (idiopathic pulmonary fibrosis, sarcoidosis and primary pulmonary hypertension); elevated pulmonary arterial pressure; and higher body mass index.^{2,33–38} Ongoing studies examining PGD mechanisms could lead to advances in the prevention or early treatment of PGD in patients with an increased risk.

Donor and recipient matching

Donor–recipient matching in LTx is usually directed by blood group (identity or compatibility) and predicted total lung capacity (pTLC) based on height, age and gender. Other donor–recipient characteristics, such as cytomegalovirus serology (+/–), gender (male/female) and age, are often ignored. The impact of donor-recipient mismatch for all these variables and their combinations on PGD occurrence has not been well investigated.³⁹

In several (single-center, multicenter, registry) studies reported by Eberlein et al, a clear correlation was found between lung size mismatch and PGD. In all studies, recipients of undersized organs (donor/recipient pTLC ≤ 1.0) had an increased PGD risk, whereas those with oversized lungs (donor/recipient pTLC > 1.0) had a reduced risk.^{40–42} This distinction was most apparent in patients without chronic obstructive pulmonary disease (COPD).⁴⁰ In addition, the post-transplant TV should be appropriate for the donor pTLC and not the recipient pTLC; otherwise, ventilation-induced lung injury with capillary leak may lead to clinical PGD.⁴³ Moreover, the potential for detrimental hyperinflation with application of negative pleural pressure to undersized lung grafts has been described.⁴⁴ On the other hand, delayed chest closure is advised in recipients with severely oversized donor lungs to avoid hemodynamic disturbances by graft compression of the heart early after LTx.⁴⁵ An oversized allograft was associated with improved post-transplant survival for idiopathic pulmonary arterial hypertension.⁴⁶ Size mismatch was also associated with long-term pulmonary allograft function and BOS, in favor

of recipients with oversized lungs.⁴⁷ In summary, incorporating the pTLC ratio into the method of lung allocation and post-transplant management could improve outcomes after LTx.

Donor–recipient matching for gender in organ transplantation is usually not considered important and so all 4 gender combinations may be possible. Few reports, mostly single-center studies with limited sample size focused on the impact of gender matching on outcome after LTx, but not specifically in relation to PGD. These studies reported a conflicting impact of gender mismatching on early and late survival and BOS.^{39,48–55} In the largest study ($n = 9,651$) using ISHLT Registry data, Sato et al found that the combination of female donor to male recipient was associated with a higher 90-day mortality and lower overall survival, whereas female donor to female recipient was associated with the best overall survival, after adjusting for size mismatch and diagnosis.⁵¹ Three other studies with high patient numbers reported similar conclusions.^{39,49,53} It remains an open question whether donor-recipient gender mismatch is an independent risk factor for early mortality after LTx,⁵¹ or whether worse survival in the female donor/male recipient combination is confounded by size mismatch, with more frequent use of undersized female donor lungs into male recipients.^{39,40,53}

In addition to matching of demographic variables, greater attention should be given to balancing known PGD risk factors in donor and recipient. Caution and clinical judgment are still needed when matching ECD lungs to high-risk recipients, especially recipients with pulmonary hypertension.^{9,56,57} Given the complexity of the interaction of multiple donor and recipient risk factors, algorithms that aid donor-recipient matching should be developed, with the aim of reducing the incidence of PGD.

Risk stratification of patients before LTx is important for several reasons. First, better identification of higher and lower risk recipient groups may allow the care team to better prepare for the likelihood of PGD development. Second, improved pre-operative prediction may facilitate safer expansion of the donor pool by characterizing lower risk recipient groups. Third, identification of higher risk recipient groups can facilitate clinical trials aimed at reducing PGD by investigating therapeutic interventions before and/or immediately after LTx. In this regard, a study by the Lung Transplant Outcomes Group produced valid estimates of PGD risk using readily available clinical variables.⁵⁸

Donor lung preservation and storage

No large, prospective, randomized trials related to lung preservation have been performed until recently. Thus, the evidence supporting the practice, as just described, is the best available and is primarily based on findings from laboratory research and single- or multicenter cohort studies performed over the last 5 decades.^{59–61} The most widely accepted and utilized conditions for lung flush and storage can be summarized as follows: preservation solution:

extracellular-type; flush volume: 60 ml/kg antegrade and 4 × 250 ml retrograde via the pulmonary veins; flush and storage temperature: 4°C; pulmonary artery flush pressure: <30 cmH₂O; route of flush: antegrade + retrograde; oxygen concentration before storage: FIO₂ 0.3 to 0.5; cold ischemic time: preferably <6 to 8 hours.

The following sub-sections provide updated information regarding the conditions for optimizing lung preservation and storage, including the potential role of EVLP, based on laboratory and clinical evidence since development of the 2005 ISHLT PGD definition.¹

Preservation solutions

No large, randomized trials are available to demonstrate the superiority in terms of outcome of one preservation solution over another. Four published clinical reports from different institutions have compared post-transplant outcomes with various preservation solutions. In a study using UNOS data between 2005 and 2008, Perfadex solution was found to be superior to University of Wisconsin solution in high-risk lung transplant recipients, with lung allocation scores (LAS) of >37.8 (1-year survival 81.5% vs 73.5%, respectively; $p = 0.02$).⁶² Marasco and colleagues, in a multivariable analysis with propensity score matching, compared the impact of 3 preservation solutions (Euro-Collins, Papworth and Perfadex) on outcomes in 310 lung transplant recipients.⁶³ Papworth solution was associated with significantly higher mortality, whereas Perfadex was associated with a lower PGD incidence at T48. Data from the UK Cardiothoracic Transplant Audit were analyzed for possible differences among current lung preservation techniques.⁶⁴ Between 1995 and 2003, 681 lung transplants were preserved with either Euro-Collins solution ($n = 284$), blood albumin ($n = 139$), core cooling ($n = 107$) or low-potassium dextran solution ($n = 151$). There was a significantly increased use of low-potassium dextran solution over time. Risk-adjusted survival was similar across the groups and was not affected by ischemic time. Survival rates at 3 years and freedom from death caused by PGD were highest in the low-potassium dextran group and lowest in the blood albumin group (62% vs 49%, and 95% vs 91%, respectively). The Hannover group recently reported their experience with 2 extracellular-type preservation solutions, comparing historical cohorts: Perfadex ($n = 209$) from 2002 to 2005 vs Celsior ($n = 208$) from 2005 to 2009.⁶⁵ Overall 3-year survival was comparable (66.5% vs 72.0%, respectively; $p = 0.25$), with significantly longer ischemic times in the Celsior cohort (355 ± 105 minutes vs 436 ± 139 minutes; $p < 0.001$). Patients with PGD 3 who received Perfadex had significantly lower survival rates at 1, 2 and 3 years after LTx when compared with patients who received Celsior. Freedom from BOS was also lower in the Perfadex group.⁶⁵

Flush and storage temperature

Lungs are generally stored at 4°C. Animal experiments have shown that 10°C may be superior, presumably due to

preservation of membrane sodium potassium channel function.⁶⁶ Recently, the University of Groningen group revisited the question on the best temperature for pulmonary flush and storage. In a rat lung transplant model after 24 hours of inflated storage, the authors found that room temperature flushing, followed by storage on ice, provided the best method for lung graft preservation.⁶⁷

Route of flush

Antegrade flush through the pulmonary artery has been standard practice since the first lung transplants were performed in the 1980s. A subsequent retrograde flush through the veins was found to have additive benefit,^{68,69} and hence has been adopted by most as standard practice. Its beneficial effect may be related to better preservation of the bronchial tree by flushing the bronchopulmonary collaterals and by removing small clots and debris from the pulmonary arterial tree. Gohrbandt et al compared antegrade flush only ($n = 173$) vs retrograde flush only ($n = 36$) in a group of 209 Hannover recipients with lungs preserved using Perfadex.⁷⁰ PGD 3 was comparable between groups at T0 to T48, but significantly higher in the retrograde group at T72 (2.2% vs 14.8%, respectively; $p < 0.05$). Bronchial dehiscence occurred more frequently in the retrograde-only group (0.6% vs 5.6%, respectively; not statistically significant [NS]), whereas bronchial stenosis occurred more often in the antegrade-only group (24.9% vs 13.9%, respectively [NS]). Overall survival and BOS-free survival were similar.⁷⁰

Tolerable cold ischemic time

Presently, most teams still tend to limit the ischemic times to <8 hours with extracellular-type solutions, although successful outcomes have been reported with longer times. Thabut et al, in a large French multicenter retrospective study, examined the impact of graft ischemic time on early and late outcomes in 752 patients after all types of LTx over a 12-year period.⁷¹ Mean graft ischemic time was 246 ± 96 minutes (range 50 to 660 minutes). After adjustment for 11 potential confounders, graft ischemic time was associated with early PGD (PaO₂/FIO₂ at T0 and T6) and with long-term survival in patients undergoing single or double LTx, but not in patients undergoing heart-lung transplantation. The hazard ratio for death with longer ischemic times increased sharply after 330 minutes, and these results were unaffected by the preservation fluid used (intracellular-type vs extracellular-type). In a recent study based on a large data set from UNOS, however, prolonged (>6 hours) cold ischemic time did not impact survival at 1 and 5 years after LTx and was also not a negative predictor of primary graft failure.⁷²

Using a porcine left single-LTx model, University of Toronto investigators examined the effect of a second cold ischemic period on allograft function, after an initial 10-hour period at 4°C followed by 6 hours of EVLP.⁷³ No differences in allograft parameters were observed at 4 hours

post-transplant between animals receiving lungs with a second 2-hour vs 10-hour cold ischemic period after EVLP. Oxygenation in both groups was superior to that of recipients of lungs preserved at 4°C for 24 hours without EVLP. These findings, if confirmed, may help to extend lung preservation time to >18 hours and may redefine the logistics of transplantation.⁷⁴

Cold static storage vs warm preservation with EVLP

Cold pulmonary flush and static storage is currently the clinical standard for lung preservation, with the intention of lung protection by slowing cell metabolism to prevent cell death and organ deterioration.^{59–61} Currently, normothermic dynamic preservation with the aid of ex-vivo perfusion is being investigated for all solid organs, including the lung.^{75–79}

The modern success of prolonged (12 hours) EVLP without edema formation is in part due to improvements in technology and in part due to the use of a buffered, extracellular solution with an optimal colloid osmotic pressure.⁸⁰ In a porcine LTx study at the University of Toronto, ongoing lung injury after 12 hours of cold storage was prevented when followed by 12-hour normothermic EVLP compared with a control group with 24 hours of cold storage.⁸¹ A prospective, international, multicenter, non-inferiority clinical trial randomized 320 bilateral, standard-criteria donor lung recipients between cold storage and immediate normothermic portable ex-vivo machine preservation with OCS Lung (Transmedics, Inc., Andover, MA) (Inspire trial: ClinicalTrials.gov NCT 01630434).⁸² The final results were presented at the 17th Congress of the European Society for Organ Transplantation, Brussels, Belgium.⁸³ In an effort to answer the question as to whether *all* donor lungs should be treated with a period of normothermic EVLP, a prospective, single-center clinical trial was conducted by the Vienna lung transplant team. They randomized 80 patients to transplantation directly after standard cold preservation with Perfadex or cold preservation plus 4 hours with normothermic in-hospital EVLP, using the Toronto technique. There were no statistically significant outcome differences demonstrable between groups.⁸⁴

Ex-vivo methods to evaluate and recondition lungs

Unrecognized injury to the donor lung may become apparent during EVLP, allowing irrecoverably injured donor lungs with deteriorating graft function during EVLP to be declined before transplantation, thus avoiding the risk of severe PGD in the recipient. The first clinical EVLP cases were reported by Steen et al from the University of Lund, both for assessment of an uncontrolled DCD lung⁸⁵ as well as for reconditioning of an unacceptable DBD lung.⁸⁶ Since then, others have reported case series with good outcomes in ECD lung recipients after EVLP resuscitation.^{17,87–101} The overall lung yield after EVLP across all reported series is around 80%.⁷⁹ Studies have suggested a lower rate of PGD

3 in recipients of initially rejected lungs undergoing EVLP compared with lungs that were grafted immediately.^{89,99}

The role of EVLP for assessment and reconditioning of questionable donor lungs is being investigated in several clinical trials.^{79,102,103} The first clinical trial was conducted by the Toronto Lung Transplant Program. In the HELP (Human Ex-Vivo Lung Perfusion) trial, high-risk lungs, which otherwise would not be used, were assessed with EVLP.^{88,89} Eighty percent of the lungs that originally did not meet acceptance criteria from both DBDs and DCDs were ultimately transplanted after EVLP and resulted in equivalent recipient outcomes compared with those of contemporary standard control donor (SCD) lungs. Rates of PGD 3 at 72 hours after transplantation were reported to be low (2% in EVLP lungs vs 8.5% in SCD lungs).⁸⁹ Other multicenter trials of EVLP for questionable lungs (NOVEL, DEVELOP, EXPAND, PERFUSIX) are ongoing and final reports are pending.^{104–108}

Besides normothermic preservation and evaluation, EVLP holds great promise for treating damaged donor lungs. Ongoing research is investigating whether lungs injured by a variety of mechanisms (brain death, contusion, aspiration, infection, edema, atelectasis) can be repaired so that some of these can become transplantable. Diagnostic strategies and targeted therapies for ex-vivo delivery will need to be developed for each of these types of injury. Potential strategies include controlled perfusion and ventilation, inhaled drugs and gases, perfusate additives and gene and cell therapy.^{79,103} Thus, EVLP has the potential to reduce the incidence of PGD 3 by identifying and reversing insults in the donor organ and limiting ischemia.

Lung implantation and reperfusion techniques

The ultimate outcome of a lung transplant operation has contributions related to: (i) the donor and the cause of death and associated injuries; (ii) the lung retrieval and preservation technique; (iii) the implantation procedure and finally; (iv) the recipient. Each of these phases of the transplant procedure will need to be optimized to reduce the incidence of PGD.

Cooling during implantation

Under normothermic ischemic and anoxic conditions, lung tissue cells deteriorate very rapidly, and lung adenosine triphosphate (ATP) stores are essentially depleted in 20 minutes of warm ischemia and anoxia. Cold flush preservation has been the cornerstone of organ preservation that made transplantation possible. Stored in the inflated state, the lung has a unique preservation advantage of having a source of oxygen—hence it is in a state of hypothermic aerobic preservation.^{109,110} Hypothermic static flush preservation with low-potassium dextran solution (Perfadex) has utilized this strategy to provide safe, reliable extended lung preservation.

Normothermic EVLP techniques provide a unique opportunity to bring the lung back to normal temperature

to evaluate and treat the lung, but these approaches add a degree of complexity. Once the lung is taken out of the storage phase and implanted into the patient, it is atelectatic and warm and is once again vulnerable to warm, ischemic and anoxic injury. Hence, for this phase of LTx it is still recommended that the lung be kept cool. This is achieved by a terminal flush cooling after EVLP and placing the lung on a water-cooled cooling jacket in the chest for the implantation procedure. For standard cold flush-preserved lungs, the lungs are taken out of the cooler and placed on the cooling jacket. The underlying principle is centered on the use of cold when it is needed for protection and warm when it is needed for assessment and treatment.

Intra-operative extracorporeal support

Extracorporeal support is used selectively to perform LTx. The majority of LTx operations can be performed without the use of extracorporeal support as a single-lung transplant or sequential bilateral lung transplant technique. However, if the patient has significant pulmonary hypertension or becomes unstable during the procedure (either hypotensive or hypoxic), then one should not hesitate to use extracorporeal support. Some have advocated routine use of cardiopulmonary bypass (CPB) for all transplants, but this needs to be considered in the balance of inflammation and anti-coagulation-related morbidity vs benefit of technical ease in patients that can be transplanted without CPB.^{111,112} The recent trend has been to use an extracorporeal lung support (ECLS) with ECMO circuit instead of conventional CPB, as it allows for less anti-coagulation, less coagulopathy and decreased use of blood products.^{113–118} In the Lung Transplant Outcomes Group study, CPB was associated with higher rates of PGD; however, planned vs emergent CPB use was not prospectively recorded.²

Blood and blood product use

The Transfusion-related Acute Lung Injury (TRALI) is a well-described phenomenon of inflammatory lung injury and an ARDS picture after significant transfusion of blood products. The already-injured lung (with ischemia-reperfusion injury) is vulnerable to this as an additional “hit,” and therefore it is advisable to avoid blood transfusion wherever possible.² Furthermore, use of the cardiotomy sucker in standard CPB is also not advisable as the activated blood likely contributes to further lung injury as an extrapolation of the aforementioned concept. Use of a cell saver (also necessary for ECMO circuits) is the favored way to auto-transfuse shed blood as concentrated and washed red cells are returned to the patient.

Methods to control reperfusion

The endothelial and epithelial cells in hypothermically preserved lungs have stiff cell membranes and the rapid reintroduction of pulmonary artery (PA) blood flow can inflict a significant shear stress injury to the lung. Studies

have demonstrated that gradual reintroduction of blood flow over a period of 10 minutes—by slowly releasing the PA clamp, can significantly improve graft function.^{119,120} If the transplant is performed on ECLS support, it is important to maintain perfusion (some ejection by the right ventricle) and ventilation to the newly implanted lung while the second lung is implanted. This avoids adding warm atelectatic ischemic injury and allows the lung to begin its recovery phase in a setting of protective ventilation and protective (low-pressure, low-flow) perfusion. Studies have reported on reperfusion with leukocyte-filtered blood^{121,122} and on adding pharmacologic agents to the initial perfusate of the implanted lung.¹²³ Ideally, as we gain better understanding of the mechanisms of the reperfusion state, these types of interventions can be applied to specifically target known components of the injury.

Principles of protective ventilation

Ventilation of the newly implanted lung should be protective. This includes keeping the FIO₂ low during the early reperfusion period—in the range of 0.21 to 0.5. Ventilation control is usually set at the lowest pressure controlled setting to achieve a reasonable TV and a PEEP of 5 to 8 cmH₂O to assist with gradual recruitment of alveoli.¹²⁴

Prophylactic post-operative ECLS (ECMO)

ECLS has traditionally been used as an advanced support measure for severe PGD in the early post-transplant phase (see next subsection). It is generally accepted that earlier institution of ECLS leads to improved salvage rates rather than delayed implementation.^{125,126} As an extension of this, some centers have advanced the concept further to leave patients on ECLS for a period of recovery (1 to 3 days or so) when early graft dysfunction is manifest in the operating room, or even to leave patients on ECLS “prophylactically” in high-risk recipient cases, such as those with primary pulmonary hypertension.^{127,128} Using ECLS as a “prevention of PGD” strategy in this fashion is somewhat surgeon- and center-specific and further study is required to determine the threshold at which ECLS could be applied as a “prophylactic” strategy to protect the newly reperfused lung in prevention of severe PGD.

Treatment of PGD

General principles of post-transplant PGD management

There is no consensus on the treatment of PGD, primarily due to a lack of appropriately powered clinical studies on the topic. However, there are many similarities between PGD and ARDS, as they are both characterized by severe hypoxemia and radiographic evidence of diffuse alveolar infiltrates.^{1,129} Most transplant centers tailor their therapies for PGD based on extrapolated treatments of ARDS.^{2,130}

As with ARDS, the overall goal in management of patients with PGD is to minimize oxygen toxicity and to prevent the volutrauma and barotrauma associated with mechanical ventilation.¹³¹ Although the mainstay of therapy remains supportive care in the majority of cases of PGD, there have been data published on utilizing ECMO in patients with severe PGD to enable more effective supportive care with less toxicity to the lung allograft.^{132–134}

Ventilator management

There are no clinical studies that have evaluated the various mechanical ventilation modalities in patients with PGD, but, given the radiographic and clinical similarities to ARDS,²⁶ many centers use lung protective ventilation (also known as low TV ventilation) with PEEP as a mainstay of therapy.¹³⁵

Fluid management

In general, fluid restriction should be used in conjunction with lung-protective ventilation in patients with PGD. However, it is important to maintain adequate perfusion so there is less systemic cytokine release. Judicious diuresis and fluid control can be used to minimize systemic perturbations in hemodynamics while avoiding worsening capillary leak and pulmonary edema. This is typically achieved by optimizing blood counts via transfusions and using systemic inotropes and/or vasopressors. Of note, the optimal hemoglobin level post-transplant has not been determined.⁴

Pulmonary vasodilators

Nitric oxide

Preservation and reperfusion of donor lungs markedly reduces nitric oxide (NO) availability.¹³⁶ Ischemia–reperfusion injury after LTx, characterized by increased capillary permeability and the development of non-cardiogenic pulmonary edema, is thought to be a hypoxic injury resulting in alterations in inflammatory mediators followed by a decline in endogenous NO. Several experimental animal model studies have shown that administration of NO to the lung allograft results in decreased pulmonary vascular resistance as well as neutrophil adhesion and platelet aggregation.^{137,138} Further experimental animal studies showed improved lung allograft function with NO treatment.^{139,140} However, small, randomized clinical trials failed to show that prophylactic inhaled NO (iNO) had an impact in prevention of PGD.^{141–143} A recent systematic review and meta-analysis for recipient-related clinical risk factors for PGD also found no significant association between the use of intra-operative iNO and development of PGD.¹⁴⁴ However, none of the trials performed have been powered to detect potentially small, but real, differences in PGD.

Similar to ARDS treatment,¹⁴⁵ NO may be useful in the treatment of established PGD by reducing pulmonary

vasoconstriction and enhancing ventilation-perfusion matching. Although case reports exist with conflicting evidence regarding clinical outcomes, there continues to be a lack of randomized clinical studies evaluating the use of NO in the treatment of PGD after LTx.^{146–149}

At this time, we cannot recommend routine prophylactic use of NO for the prevention of PGD. However, NO may be used selectively in patients with established PGD 3 showing severe hypoxemia and elevated pulmonary artery pressures as part of overall treatment program.

Prostaglandins

Prostaglandins such as PGI₂ (epoprostenol) and PGE₁ (alprostadil) play a significant role in pulmonary vasodilation and inhibition of inflammatory events such as disruption of the alveolar-capillary barrier, leukocyte adhesion and platelet aggregation.¹⁵⁰ These effects are mediated through cyclic adenosine monophosphate (cAMP) pathways that are disrupted with the ischemia-reperfusion injury in LTx recipients. Similar to NO, administration of prostaglandins has been shown to reduce pulmonary vascular resistance and improve oxygenation.^{151–153} During transplant, administration of inhaled prostaglandins before organ procurement or after implantation has resulted in decreased inflammatory cytokines, decreased pulmonary edema, decreased pulmonary artery pressure and central venous pressure, and improved cardiac index and mixed venous oxygen saturation.^{153–155} However, no studies to date have evaluated the effects of these changes on clinical PGD.

At this time, there is insufficient data to recommend the use of routine inhaled prostaglandins for the prevention of PGD after LTx, although the use of inhaled prostaglandins for severe hypoxemia or elevated pulmonary artery pressures may prove to be a useful adjunct, similar to inhaled NO.

Novel therapies for prevention and treatment of PGD

Novel therapies for PGD have focused on prevention over treatment and include surfactant, complement inhibition, platelet-activating factor antagonists, platelet and neutrophil traps, stem cells and plasmapheresis. Human studies evaluating their role in the prevention and treatment of PGD are listed in [Table 1](#). More details on these novel therapies are given in [Appendix 2](#).

ECMO

A subset of patients with PGD 3 has the potential to benefit from post-transplant ECMO. In most cases of PGD, optimization of conventional ventilator support allows for recovery of the injured lung. However, mechanical ventilation alone cannot achieve sufficient gas exchange in some patients with severe PGD. In such cases, the therapeutic interventions mentioned earlier may be utilized. If these therapies fail, ECMO is recommended. The general

Table 1 Summary of Human Studies Evaluating Novel Interventions for Prevention and Treatment of PGD After Lung Transplantation

Therapy	Design	Results	Author and reference no.
Surfactant: endobronchial instillation of 20 mg/kg before reperfusion	RCT: 42 recipients	Lower PGD, earlier extubation	Amital et al ¹⁸⁹
Surfactant: endobronchial instillation 3 to 7 days post-Tx	Case series: 5 patients	Improved P/F ratio in the setting of severe PGD	Amital et al ¹⁹¹
Complement inhibition: TP-10 (10 mg/kg) before reperfusion	RCT: 59 patients	No significant difference in time on vent or ICU stay	Keshavjee et al ¹⁹³
Platelet-activating factor antagonist: 2 mg/kg and 10 mg/kg	RCT: 24 patients	Short-lived improved P/F ratio < 8 hours after Tx	Wittwer et al ²¹⁰

ICU, intensive care unit; P/F, PaO₂ to FIO₂ ratio; PGD, primary graft dysfunction; RCT, randomized, controlled trial; Tx, transplant.

indication for ECMO in PGD is severe hypoxemia (PO₂/FIO₂ <100 mm Hg) not responsive to pulmonary vasodilation, with or without hypercapnia, acidosis and right ventricular dysfunction.

One of the major concerns when using ECMO in the post-transplant period has been the high incidence of complications, such as bleeding, vascular injury and neurologic deficits. According to the ARDS literature, the incidence of such complications has dramatically decreased in recent years. The main reasons for this decrease are: (1) improved device technology leading to less blood trauma and the requirement of lower anti-coagulation parameters (activated clotting time 160 to 180 seconds or activated partial thromboplastin time 1.5 to 2 times normal); (2) earlier implementation of ECMO so patients are not in multiorgan failure; and (3) broad use of venous-venous (V-V) ECMO instead of venous-arterial (V-A) ECMO. V-V ECMO can properly support the majority of patients with severe PGD, even in the setting of hemodynamic compromise. Correction of hypoxemia and acidosis with V-V ECMO and pulmonary vasodilation due to oxygenated blood perfusing the lungs often leads to rapid hemodynamic improvement, which nullifies the need for V-A ECMO. One exception to this generalization is in patients with primary pulmonary hypertension, where V-A or V-VA hybrid ECMO is often extended to the post-transplant period to protect the left ventricle from overflow and subsequent cardiogenic pulmonary edema.^{127,128}

A topic of continued controversy is whether V-A ECMO can better protect the injured lungs by offloading the pulmonary circulation. Although this assumption is physiologically sound, one needs to balance the risks and benefits of such approach vs V-V ECMO. Some data also support V-V ECMO as a strategy to decrease pulmonary artery pressures (PAPs) with an average PAP decrease of 20 mm Hg after initiation.¹⁵⁶ Reduction of pulmonary perfusion with V-A ECMO may also lead to increased incidence of bronchial complications, as bronchial vascularization is dependent on pulmonary flow in the early post-transplant period. Other concerns with V-A ECMO include increased vascular complications from the arterial puncture site, increased neurologic complications, and need for anti-coagulation.

Exemplifying clinical practice, Bermudez and colleagues reported on a large proportion of V-A ECMO use for PGD, with

survival rates of 56%, 40% and 25% at 30 days, 1 year and 5 years, respectively.¹³³ In contrast, Hartwig and colleagues used V-V ECMO for PGD in 6% of their transplant population with survival being substantially better than in previous reports: 30 days, 82%; 1 year, 64%; and 5 years, 49%.¹³⁴ Thus, V-V ECMO has been the growing mode of choice for extracorporeal life support in patients with severe PGD.

Taken together, these findings indicate that post-transplant ECMO is useful for supporting patients through PGD, but this population still has significantly lower long-term survival than patients without severe PGD.

Retransplantation

Less than 5% of all lung transplants performed are retransplants.^{157,158} Despite improvements in overall outcomes, repeat LTx demonstrates worse overall survival than primary LTx. Early "re-do" LTx survival remains particularly hazardous, and this should remain an option of last resort for the treatment of PGD. Ideally, a patient with PGD can be managed with the more conservative measures, as described earlier in this report. This would include mechanical support such as V-V ECMO, which has proven to be an invaluable and safe mechanism for bridging patients through severe PGD. If these other means of treatment and support prove unsuccessful, then the option to retransplant the patient may be entertained. The clinical team should not take this decision lightly as it presents a tremendous challenge to resources and likely endangers another pulmonary allograft that may be more appropriately utilized in another recipient.

The ISHLT Registry data indicate that survival after repeat LTx remains well below that of primary LTx.¹⁵⁷ However, the median survival after retransplantation has improved over time and, in the most recent era, has reached 3.0 years. A similar result can be seen in the UNOS registry.¹⁵⁹ Some of this may be due to improvements in best available care, but some of the improvements may also be secondary to implementation of the LAS in the United States and elsewhere, which more quickly triages available allografts to these patients over less ill ones. What is clear when analyzing both data sets is that the interval between

primary transplantation and retransplantation is strongly associated with survival. In the ISHLT Registry, those patients retransplanted within 1 month of the primary transplant had a median survival of <6 months. Similarly, early retransplant, <90 days after primary transplant in the USA experience, showed a hazard ratio (HR) = 2.4 compared with late retransplant and an HR = 3.1 compared with primary transplant. Likewise, in the USA, those patients specifically retransplanted for PGD had worse survival compared with those transplanted for BOS (HR = 1.63, 95% confidence interval 1.11 to 2.38), even since implementation of the LAS. Although the data would suggest that peak allograft function is attenuated in patients with severe PGD,¹³⁴ a better resource utilization strategy may be to support patients with severe PGD through the process and then perform late retransplant if necessary, when overall survival is expected to be better.

Conclusions

Dysfunction of the pulmonary allograft during the first 72 hours after LTx may be the end result of several physiologic and biochemical insults that occur during the transplantation process, namely: in the donor before and after death; during flush preservation and storage; during implantation; and after reperfusion in a specific recipient. For the best possible outcome, each of these phases of the transplant procedure will need to be optimized. There has been significant progress in the past decade in identifying donor and recipient risk factors for PGD.

Thankfully, in most patients, PGD is of mild to moderate severity, and can be managed with standard supportive therapy in the intensive care unit. In some patients, however, PGD can be severe. Many findings have been published on utilizing ECMO early in the post-transplant course to support these patients who are at significant risk for early death.

As we learn more about the mechanisms of this injury, new strategies need to be developed to specifically ameliorate each component of the injury process. Multi-center clinical trials would be helpful in determining the best management for LTx recipients with PGD. The goal of the LTx community is to reduce the risk for early death and improve early- and long-term outcome after LTx for patients afflicted with PGD at all levels of severity.

Disclosure statement

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Appendix 1. : ISHLT PGD Working Group IV

Members of the ISHLT PGD Working Group IV include: Marcelo Cypel, R. Duane Davis, Matthew G. Hartwig, Don Hayes, Jr, Steve Ivulich, Marshall I. Hertz, Shaf Keshavjee, Jasleen Kukreja, Erika Lease, Gabriel Loor, Olaf Mercier, Luca Paoletti, Jasvir Parmar, Reinaldo Rampolla, Dirk Van Raemdonck, Rajat Walia, and Keith Wille.

Appendix 2. : Novel therapies for prevention and treatment of PGD

1. Surfactant

Pulmonary surfactant is a heterogeneous, surface-active lipoprotein complex that is composed of 90% lipids (65% phosphatidylcholine) and approximately 10% serum-derived proteins, with the latter representing the 4 surfactant-associated proteins (SP-A, SP-B, SP-C and SP-D).^{160,161} After synthesis by Type II pneumocytes, surfactant is secreted into the alveolar space where it forms a stable monolayer, resulting in reduced surface tension of the alveoli and stabilization during end-expiration, prevention of atelectasis and alveolar edema and an optimal surface area for gas exchange.^{160,161} Clinical and experimental studies have demonstrated that ischemia, cold storage and reperfusion associated with LTx influences surfactant composition and function,^{162–167} including alterations in the surfactant aggregate ratio with decreases in phosphatidylcholine and phosphatidylglycerol. These alterations correlate with a reduction in pulmonary compliance, resulting in alveolar collapse, ventilation-perfusion mismatch, pulmonary edema and decreased oxygenation.^{168–170} Moreover, preliminary experimental studies and clinical experience with exogenous administration of surfactant have shown partial mitigation of these complications.^{162–165,171–182}

Although exogenous surfactant appeared to be beneficial in LTx in early studies, critical questions remain that include dosing and timing of administration (e.g., at procurement vs reperfusion, or both). In an important experimental model, attenuation of lung ischemia-reperfusion injury by pre-ischemic exogenous surfactant was described through stabilizing and increasing the active endogenous intra-alveolar surfactant pool.¹⁸³ Donor lung pre-treatment with an SP-A-free surfactant agent also maintained serum NO and reduced hemodynamic disturbances, while better preserving alveolar integrity in a porcine LTx model.¹⁷⁰ Likewise, in another experimental model, treatment with surfactant before lung reperfusion resulted in improved lung compliance, improved oxygenation, decreased protein leakage and enhanced survival.^{184,185} Again, when administered before ischemia-reperfusion injury, intratracheal surfactant application significantly reduced intra-alveolar edema and prevented atelectasis, whereas peribronchovascular edema increased and alveolar Type II cells' morphologic alterations were not influenced by surfactant treatment,¹⁸⁶ suggesting the benefit of exogenous surfactant is related to intra-alveolar activity. In a murine model with

exogenous surfactant being effective in both prevention and treatment of lung ischemia-reperfusion injury, Wittwer et al¹⁸⁷ evaluated its application at the time of flush preservation, after 4-hour ischemia and during reperfusion and found that donor lung pre-treatment with endobronchial surfactant provided optimal preservation quality compared with post-ischemic application or during reperfusion.

Several groups have undertaken research examining the benefit of exogenous surfactant in human LTx. The Hannover group performed a single-center, prospective, randomized trial in which surfactant was instilled by bronchoscopy into donor lungs before retrieval; the surfactant study group had higher phospholipids in bronchoalveolar lavage fluid and improved surfactant function based on enhanced small-to-large aggregate ratio.¹⁸⁸ Clinically, the patients given surfactant had a significantly higher pulmonary function 1 month after transplant, but this difference disappeared by the end of the first post-transplant year.¹⁸⁸ In another single-center, prospective, randomized study,¹⁸⁹ surfactant was delivered through a bronchoscope after establishment of bronchial anastomosis and demonstrated improved oxygenation, fewer radiographic abnormalities, lower PGD grade, reduced severe PGD rates, earlier extubation and shorter intensive care unit length of stay, along with better short-term pulmonary function outcomes. As a salvage therapy after ischemia-reperfusion injury or development of PGD in lung transplant recipients, 2 case series successfully used exogenous surfactant with significant improvement in oxygenation and resolution of radiologic infiltrates and showed excellent short-term graft function.^{190,191}

For now, exogenous surfactant therapy remains as a promising therapeutic option for lung ischemia-reperfusion injury in LTx, with associated improvement in oxygenation, prevention of PGD, and optimal short-term clinical outcomes in single-center studies and case series. Although there is a lack of rigorous prospective, randomized studies, the available data suggest surfactant may best be used in a preventive, as opposed to therapeutic, manner.

2. Complement inhibition

With activation of the complement system having an important role in mediating reperfusion injury after LTx, early interference of this pathway has been examined as a potential therapeutic target to reduce lung reperfusion injury in LTx. Complement activation accelerates tissue injury directly by complement factors or indirectly by complement-mediated polymorphonuclear neutrophil activation. In a large-animal model in which C1-esterase inhibitor was infused (half the dose given 10 minutes before and the other half 10 minutes after reperfusion), reduced lung ischemia-reperfusion injury and improved pulmonary function were observed.¹⁹² Starting in the late 1990s, experimental studies, case reports and a multicenter, randomized, double-blinded, placebo-controlled trial showed early evidence that soluble complement receptor-1 (sCR1), a complement inhibitor, may be beneficial for the treatment of PGD.^{193–197} In the

randomized, double-blinded, multicenter, placebo-controlled trial, where 28 patients received sCR1 and 31 received placebo before reperfusion, early outcomes were improved in the setting of 90% complement inhibition for 24 hours, with a return to normal activity by 72 hours.¹⁹³ Combining the complement inhibition of sCR1 with the leukocyte adhesion inhibition of selectin ligand sialyl Lewis X (sLeX) resulted in a significant reduction of reperfusion injury in experimental models, with no effect on graft rejection.^{198,199} More recently, the role of complement-mediated microvascular injury in chronic lung allograft rejection has been under investigation.²⁰⁰ In kidney transplantation, targeting complement activation in the donor after brain death improved short-term renal function after transplantation in recipients.²⁰¹ With clear evidence that complement inhibition improves early outcomes, further research is needed to determine optimal techniques of administration of these potential therapies at time of reperfusion in LTx in order to prevent PGD and potentially improve long-term outcomes.

3. Platelet-activating factor antagonists

Platelet-activating factor (PAF) is a phospholipid that is released during ischemic lung injury, so PAF antagonists were initially described as a potential adjunct for lung preservation in the early 1990s.^{202–208} A PAF antagonist combined with an endothelin antagonist in an experimental model showed superior post-transplant graft function 24 hours after reperfusion compared with no treatment and each individual agent, suggesting a synergistic role.²⁰⁹ A 2001 study in humans demonstrated significant improvement in alveolar-arterial oxygen differences for the first 12 hours after reperfusion and better chest X-ray score in randomly assigned patients who received low-dose ($n = 8$) and high-dose ($n = 8$) PAF antagonist in the flush solution before reperfusion, when compared with a control group ($n = 8$).²¹⁰ The investigators reported a clear benefit of the PAF antagonist in the early post-ischemic period, but the distinction dissipated after 32 hours.²¹⁰ In 1994, a single-center, randomized, double-blind trial on 29 kidney transplant recipients showed significantly less primary graft failure in donor organs treated with a PAF antagonist.²¹¹ More recently, anti-PAF attenuated leukocyte adhesion response in an experimental model of bowel ischemia with application in small bowel transplantation resulted in improved sub-mucosal capillary flow and reduced tissue injury.²¹² Although investigations on the role of PAF antagonists in LTx are not as common as they were 2 decades ago, there is enough evidence suggesting they may be useful as adjunct therapy during the early post-transplant period and thus may assist in PGD treatment. Further research is needed to better define the role of PAF antagonists in LTx.

4. Platelets and neutrophil extracellular traps

Platelets may represent a target for therapy as previous studies have highlighted a potential role in lung injury.²¹³

More recently, platelet depletion in 2 mouse models of ARDS reduced the severity of lung injury and increased survival. Platelet deprivation post-operatively may not be feasible, but this effect could also be reproduced by pre-treatment with aspirin.^{214–216} Interest in pre-emptive treatment with aspirin in ARDS is being explored in high-risk pre-operative patients.²¹⁵ This may potentially be translatable to LTx recipients as a preventive or therapeutic strategy.

The presence of neutrophil extracellular traps (NETs) is thought to activate epithelial cells and increase the aggregation of platelets. This observation was confirmed in both an experimental mouse model and a human study. Targeting these NETs with DNase alone and in combination with aspirin decreased the degree of lung injury in both models.²¹⁷ One possible mechanistic explanation is the observation that delayed neutrophil apoptosis, a feature of ARDS, is decreased by aspirin, which allows for resolution of the persisting inflammation.²¹⁸

5. Stem cells

Stem cells are considered to have potent anti-inflammatory properties, through local paracrine mechanisms, as well the tantalizing potential for local regeneration. Stem cells also have the ability to donate functioning mitochondria to injured cells. Multiple animal studies in ARDS models have demonstrated an increase in the anti-inflammatory cytokines (interleukin-8 and interleukin-1 receptor antagonists), thus ameliorating inflammation. A small-scale human EVLP study involving 4 discarded donor lungs in a protracted ischemic model suggested improvement in the inflammatory profile and histology after intratracheal administration of multipotent adult progenitor cells.^{219–221} Interest in this area has grown significantly and a critical care medicine society has proposed a Phase I study, the STem Cells for ARDS Trial (START), to examine the feasibility of treatment with stem cells.²²²

6. Plasmapheresis

The presence of non-HLA antibodies has been linked to development of both PGD and BOS, particularly in cystic fibrosis and idiopathic pulmonary fibrosis patients. In a single-center retrospective study, the presence of K $_{\alpha 1}$ tubulin and collagen V auto-antibodies was associated with a significantly higher risk of PGD (88% vs 54%, $p < 0.05$) compared with control groups.²²³ The possibility of reducing these antibodies with plasmapheresis may provide a novel target for at-risk populations.

7. Future developments

The intensity of injury generated by an episode of PGD suggests that multiple pathways are synchronously activated at the time of lung reperfusion. The Lung Transplant Outcomes Group has identified a series of potential therapeutic targets related to oxidant stress that were

statistically related to PGD, including donor NADPH oxidase 3 (NOX3), glutathione peroxidase (GPX1) and nuclear factor (NRF-2). The GPX1 association included 3 individual loci (p -values between 0.006 and 0.049) and the NRF-2 (NFE2L2) association included 2 loci ($p = 0.03$ and 0.05).²²⁴ Polymorphisms in the interleukin-17 and interleukin-23 receptors of the recipients were associated with an increased incidence of PGD.²²⁵ Further elucidating the affected pathways in PGD may lead to novel therapeutic interventions.

Unfortunately, 2 large, randomized, controlled trials examining the use of β_2 -agonists and statins in ARDS failed to demonstrate a benefit from either therapy.^{226,227} However, it is of interest to note from observational data from the ISHLT Registry that patients on statins post-LTx have a better outcome than those not on statins.²²⁸ Other novel therapies, including renin-angiotensin axis blockers, peroxisome proliferator agonist receptor ligands, curcumin and inhaled heparin, are currently being considered. These approaches have some supporting data in animal models, but have limited clinical data.²²⁹

References

- Christie JD, Carby M, Bag R, et al. Report of the ISHLT Working Group on primary lung graft dysfunction part II: definition. A consensus statement of the International Society for Heart and Lung Transplantation. *J Heart Lung Transplant* 2005;24:1454-9.
- Diamond JM, Lee JC, Kawut SM, et al. Clinical risk factors for primary graft dysfunction after lung transplantation. *Am J Respir Crit Care Med* 2013;187:527-34.
- Shah RJ, Bellamy SL, Localio AR, et al. A panel of lung injury biomarkers enhances the definition of primary graft dysfunction (PGD) after lung transplantation. *J Heart Lung Transplant* 2012; 31:942-9.
- Shargall Y, Guenther G, Ahya VN, et al. Report of the ISHLT working group on primary graft dysfunction part VI: treatment. *J Heart Lung Transplant* 2005;24:1489-500.
- Van Raemdonck D, Neyrinck A, Verleden GM, et al. Donor selection and management. *Proc Am Thorac Soc* 2009;6:28-38.
- Snell GI, Westall GP. Selection and management of the lung donor. *Clin Chest Med* 2011;32:223-32.
- Orens JB, Boehler A, de Perrot M, et al. A review of lung transplant donor acceptability criteria. *J Heart Lung Transplant* 2003;22: 1183-200.
- Somers J, Ruttens D, Verleden SE, et al. A decade of extended-criteria lung donors in a single center: was it justified? *Transpl Int* 2015;28:170-9.
- Pierre AF, Sekine Y, Hutcheon MA, et al. Marginal donor lungs: a reassessment. *J Thorac Cardiovasc Surg* 2002;123:421-7.
- Botha P, Trivedi D, Weir CJ, et al. Extended donor criteria in lung transplantation: impact on organ allocation. *J Thorac Cardiovasc Surg* 2006;131:1154-60.
- Sommer W, Ius F, Salman J, et al. Survival and spirometry outcomes after lung transplantation from donors aged 70 years and older. *J Heart Lung Transplant* 2015;34:1325-33.
- Baldwin MR, Peterson ER, Easthausen I, et al. Donor age and early graft failure after lung transplantation: a cohort study. *Am J Transplant* 2013;13:2685-95.
- Bonser RS, Taylor R, Collett D, et al. Effect of donor smoking on survival after lung transplantation: a cohort study of a prospective registry. *Lancet* 2012;380:747-55.
- Cypel M, Levvey B, Van Raemdonck D, et al. International Society for Heart and Lung Transplantation donation after circulatory death registry report. *J Heart Lung Transplant* 2015;34:1278-82.

15. Krutsinger D, Reed RM, Blevins A, et al. Lung transplantation from donation after cardiocirculatory death: a systematic review and meta-analysis. *J Heart Lung Transplant* 2015;34:675-84.
16. Gomez-de-Antonio D, Campo-Cañaverl JL, Crowley S, et al. Clinical lung transplantation from uncontrolled non-heart-beating donors revisited. *J Heart Lung Transplant* 2012;31:349-53.
17. Warnecke G, Moradiellos J, Tudorache I, et al. Normothermic perfusion of donor lungs for preservation and assessment with the Organ Care System Lung before bilateral transplantation: a pilot study of 12 patients. *Lancet* 2012;380:1851-8.
18. Malinoski DJ, Daly MC, Patel MS, et al. Achieving donor management goals before deceased donor procurement is associated with more organs transplanted per donor. *J Trauma* 2011;71:990-5.
19. Franklin GA, Smith JW, Daugherty W, et al. Incremental increases in organ retrieval after protocol driven change in an organ procurement organization: a 15-year assessment. *Am Surg* 2009;75:537-43.
20. Singbartl K, Murugan R, Kaynar AM, et al. Intensivist-led management of brain-dead donors is associated with an increase in organ recovery for transplantation. *Am J Transplant* 2011;11:1517-21.
21. MacLean A, Dunning J. The retrieval of thoracic organs: donor assessment and management. *Br Med Bull* 1997;53:829-43.
22. Angel LF, Levine DJ, Restrepo MI, et al. Impact of a lung transplantation donor-management protocol on lung donation and recipient outcomes. *Am J Respir Crit Care Med* 2006;174:710-6.
23. Noiseux N, Nguyen BK, Marsolais P, et al. Pulmonary recruitment protocol for organ donors: a new strategy to improve the rate of lung utilization. *Transplant Proc* 2009;41:3284-9.
24. Mascia L, Bosma K, Pasero D, et al. Ventilatory and hemodynamic management of potential organ donors: an observational survey. *Crit Care Med* 2006;34:321-7.
25. Mascia L, Pasero D, Slutsky AS, et al. Effect of a lung protective strategy for organ donors on eligibility and availability of lungs for transplantation: a randomized controlled trial. *JAMA* 2010;304:2620-7.
26. The Acute Respiratory Distress Syndrome Network. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. *N Engl J Med* 2000;342:1301-8.
27. Minambres E, Rodrigo E, Ballesteros MA, et al. Impact of restrictive fluid balance focused to increase lung procurement on renal function after kidney transplantation. *Nephrol Dial Transplant* 2010;25:2352-6.
28. Miñambres E, Coll E, Duerto J, et al. Effect of an intensive lung donor-management protocol on lung transplantation outcomes. *J Heart Lung Transplant* 2014;33:178-84.
29. Venkateswaran RV, Patchell VB, Wilson IC, et al. Early donor management increases the retrieval rate of lungs for transplantation. *Ann Thorac Surg* 2008;85:278-86.
30. Ware LB, Landeck M, Koyama T, et al. A randomized trial of the effects of nebulized albuterol on pulmonary edema in brain-dead organ donors. *Am J Transplant* 2014;14:621-8.
31. Weill D, Benden C, Corris PA, et al. A consensus document for the selection of lung transplant candidates: 2014—an update from the Pulmonary Transplantation Council of the International Society for Heart and Lung Transplantation. *J Heart Lung Transplant* 2015;34:1-15.
32. Barr ML, Kawut SM, Whelan TP, et al. Report of the ISHLT Working Group on Primary Lung Graft Dysfunction part IV: recipient-related risk factors and markers. *J Heart Lung Transplant* 2005;24:1468-82.
33. Whitson BA, Nath DS, Johnson AC, et al. Risk factors for primary graft dysfunction after lung transplantation. *J Thorac Cardiovasc Surg* 2006;131:73-80.
34. Kuntz CL, Hadjiliadis D, Ahya VN, et al. Risk factors for early primary graft dysfunction after lung transplantation: a registry study. *Clin Transplant* 2009;23:819-30.
35. Fang A, Studer S, Kawut SM, et al. Elevated pulmonary artery pressure is a risk factor for primary graft dysfunction following lung transplantation for idiopathic pulmonary fibrosis. *Chest* 2011;139:782-7.
36. Whelan TPM, Dunitz JM, Kelly RF, et al. Effect of preoperative pulmonary artery pressure on early survival after lung transplantation for idiopathic pulmonary fibrosis. *J Heart Lung Transplant* 2005;24:1269-74.
37. Lederer DJ, Kawut SM, Wickersham N, et al. Obesity and primary graft dysfunction after lung transplantation: the Lung Transplant Outcomes Group Obesity Study. *Am J Respir Crit Care Med* 2011;184:1055-61.
38. De Oliveira NC, Osaki S, Maloney J, et al. Lung transplant for interstitial lung disease: outcomes for single versus bilateral lung transplantation. *Interact Cardiovasc Thorac Surg* 2012;14:263-7.
39. Demir A, Coosemans W, Decaluwé H, et al. Donor-recipient matching in lung transplantation: which variables are important? *Eur J Cardiothorac Surg* 2015;47:974-83.
40. Eberlein M, Reed RM, Bolukbas S, et al. Lung size mismatch and primary graft dysfunction after bilateral lung transplantation. *J Heart Lung Transplant* 2015;34:233-40.
41. Eberlein M, Reed RM, Madaa M, et al. Donor-recipient size matching and survival after lung transplantation. A cohort study. *Ann Am Thorac Soc* 2013;10:418-25.
42. Eberlein M, Reed RM, Bolukbas S, et al. Lung size mismatch and survival after single and bilateral lung transplantation. *Ann Thorac Surg* 2013;96:457-63.
43. Dezube R, Arnaoutakis GJ, Reed RM, et al. The effect of lung-size mismatch on mechanical ventilation tidal volumes after bilateral lung transplantation. *Interact Cardiovasc Thorac Surg* 2013;16:275-81.
44. Kozower BD, Meyers BF, Ciccone AM, et al. Potential for detrimental hyperinflation after lung transplantation with application of negative pleural pressure to undersized lung grafts. *J Thorac Cardiovasc Surg* 2003;125:430-2.
45. Shigemura N, Orhan Y, Bhama JK, et al. Delayed chest closure after lung transplantation: techniques, outcomes, and strategies. *J Heart Lung Transplant* 2014;33:741-8.
46. Eberlein M, Diehl E, Bolukbas S, et al. An oversized allograft is associated with improved survival after lung transplantation for idiopathic pulmonary arterial hypertension. *J Heart Lung Transplant* 2013;32:1172-8.
47. Eberlein M, Permutt S, Chahla MF, et al. Lung size mismatch in bilateral lung transplantation is associated with allograft function and bronchiolitis obliterans syndrome. *Chest* 2012;141:451-60.
48. Roberts DH, Wain JC, Chang Y, et al. Donor-recipient gender mismatch in lung transplantation: impact on obliterative bronchiolitis and survival. *J Heart Lung Transplant* 2004;23:1252-9.
49. Thabut G, Mal H, Cerrina J, et al. Influence of donor characteristics on outcome after lung transplantation: a multicenter study. *J Heart Lung Transplant* 2005;24:1347-53.
50. Sato M, Gutierrez C, Waddell TK, et al. Donor-recipient gender mismatch in lung transplantation: impact on obliterative bronchiolitis and survival. *J Heart Lung Transplant* 2005;24:2000-1.
51. Sato M, Gutierrez C, Kaneda H, et al. The effect of gender combinations on outcome in human lung transplantation: the International Society for Heart and Lung Transplantation Registry experience. *J Heart Lung Transplant* 2006;25:634-7.
52. Miñambres E, Llorca J, Subriviola B, et al. Influence of donor-recipient gender mismatch in early outcome after lung transplantation. *Transplant Proc* 2008;40:3076-8.
53. Fessard D, Dromer C, Thumerel M, et al. Influence of gender donor-recipient combinations on survival after human lung transplantation. *Transplant Proc* 2011;43:3899-902.
54. Alvarez A, Moreno P, Illana J, et al. Influence of donor-recipient gender mismatch on graft function and survival following lung transplantation. *Interact Cardiovasc Thorac Surg* 2013;16:426-35.
55. Mangiameli G, Arame A, Boussaud V, et al. Lung transplantation in childhood and adolescence: unicentric 14-year experience with sex matching as the main prognosticator. *Eur J Cardiothorac Surg* 2016;49:810-7.
56. Moreno P, Alvarez A, Santos F, et al. Extended recipients but not extended donors are associated with poor outcomes following lung transplantation. *Eur J Cardiothorac Surg* 2014;45:1040-7.
57. Shigemura N, Horai T, Bhama JK, et al. Lung transplantation with lungs from older donors: recipient and surgical factors affect outcomes. *Transplantation* 2014;98:903-38.

58. Shah RJ, Diamond JM, Cantu E, et al. Objective estimates improve risk stratification for primary graft dysfunction after lung transplantation. *Am J Transplant* 2015;15:2188-96.
59. Van Raemdonck D. Thoracic organs: current preservation technology and future prospects; part 1: lung. *Curr Opin Organ Transplant* 2010;15:150-5.
60. Machuca TN, Cypel M, Keshavjee S. Advances in lung preservation. *Surg Clin N Am* 2013;93:1373-94.
61. Munshi L, Keshavjee S, Cypel M. Donor management and lung preservation for lung transplantation. *Lancet Respir Med* 2013;1:318-28.
62. Amaoutakis GJ, Allen JG, Merlo CA, et al. Low potassium dextran is superior to University of Wisconsin solution in high-risk lung transplant recipients. *J Heart Lung Transplant* 2010;29:1380-7.
63. Marasco SF, Bailey M, McGlade D, et al. Effect of donor preservation solution and survival in lung transplantation. *J Heart Lung Transplant* 2011;30:414-9.
64. Ganesh JS, Rogers CA, Banner NR, et al. Does the method of lung preservation influence outcome after transplantation? An analysis of 681 consecutive procedures. *J Thorac Cardiovasc Surg* 2007;134:1313-21.
65. Gohrbandt B, Simon AR, Warnecke G, et al. Lung preservation with Perfadex or Celsior in clinical transplantation: a retrospective single-center analysis of outcomes. *Transplantation* 2015;99:1933-9.
66. Wang LS, Nakamoto K, Hsieh CM, et al. Influence of temperature of flushing solution on lung preservation. *Ann Thorac Surg* 1993;55:711-5.
67. Munneke AJ, Rakhorst G, Petersen AH, et al. Flush at room temperature followed by storage on ice creates the best lung graft preservation in rats. *Transpl Int* 2013;26:751-60.
68. Varela A, Cordoba M, Serrano-Fiz S, et al. Early lung allograft function after retrograde and antegrade preservation. *J Thorac Cardiovasc Surg* 1997;114:1119-20.
69. Venuta F, Rendina EA, Bui M, et al. Preimplantation retrograde pneumoplegia in clinical lung transplantation. *J Thorac Cardiovasc Surg* 1999;118:107-14.
70. Gohrbandt B, Warnecke G, Fischer S, et al. Retrograde in situ versus antegrade pulmonary preservation in clinical lung transplantation: a single-centre experience. *Eur J Cardiothorac Surg* 2016;49:55-62.
71. Thabut G, Mal H, Cerrina J, et al. Graft ischemic time and outcome of lung transplantation: a multicenter analysis. *Am J Respir Crit Care Med* 2005;171:786-91.
72. Grimm JC, Valero V 3rd, Kilic A, et al. Association between prolonged graft ischemia and primary graft failure or survival following lung transplantation. *JAMA Surg* 2015;150:547-53.
73. Hsin MK, Iskender I, Nakajima D, et al. Extension of donor lung preservation with hypothermic storage after normothermic ex vivo lung perfusion. *J Heart Lung Transplant* 2016;35:130-6.
74. Krueger T, Machuca T, Linacre V, et al. Impact of extended cold ischemic times on the outcome of clinical lung transplantation using ex vivo lung perfusion (abstract 243). *J Heart Lung Transplant* 2014;33(suppl):S94.
75. Hosgood SA, van Heurn E, Nicholson ML. Normothermic machine perfusion of the kidney: better conditioning and repair? *Transpl Int* 2015;28:657-64.
76. Ravikumar R, Leuvenink H, Friend PJ. Normothermic liver preservation: a new paradigm? *Transpl Int* 2015;28:690-9.
77. Barlow AD, Hamed MO, Mallon DH, et al. Use of ex vivo normothermic perfusion for quality assessment of discarded human donor pancreases. *Am J Transplant* 2015;15:2475-82.
78. Messer S, Ardehali A, Tsui S. Normothermic donor heart perfusion: current clinical experience and the future. *Transpl Int* 2015;28:634-42.
79. Van Raemdonck D, Neyrinck A, Cypel M, et al. Ex-vivo lung perfusion. *Transpl Int* 2015;28:643-56.
80. Cypel M, Yeung JC, Hirayama S, et al. Technique for prolonged normothermic ex vivo lung perfusion. *J Heart Lung Transplant* 2008;27:1319-25.
81. Cypel M, Rubacha M, Yeung JC, et al. Normothermic ex vivo perfusion prevents lung injury compared to extended cold preservation for transplantation. *Am J Transplant* 2009;9:2262-9.
82. International Randomized Study of the TransMedics Organ Care System (OCS Lung) for Lung Preservation and Transplantation (Inspire). <http://clinicaltrials.gov/ct2/show/NCT01630434?term=Inspire&rank=7>. Accessed April 1, 2016.
83. Warnecke G, Van Raemdonck D, Kukreja J, et al. The Organ Care System (OCS) lung inspire international trial results (abstract OLB05). *Transplant Int* 2015;28(suppl 4):131.
84. Slama A, Schillab L, Barta M, et al. Standard donor lung procurement with normothermic ex vivo lung perfusion: A prospective randomized clinical trial. *J Heart Lung Transplant* 2017;36:744-53.
85. Steen S, Sjöberg T, Pierre L, et al. Transplantation of lungs from a non-heart-beating donor. *Lancet* 2001;357:825-9.
86. Steen S, Ingemansson R, Eriksson L, et al. First human transplantation of a nonacceptable donor lung after reconditioning ex vivo. *Ann Thorac Surg* 2007;83:2191-4.
87. Ingemansson R, Eyjolfsson A, Mared L, et al. Clinical transplantation of initially rejected donor lungs after reconditioning ex vivo. *Ann Thorac Surg* 2009;87:255-60.
88. Cypel M, Yeung J, Liu M, et al. Normothermic ex vivo lung perfusion in clinical lung transplantation. *N Engl J Med* 2011;364:1431-40.
89. Cypel M, Yeung JC, Machuca T, et al. Experience with the first 50 ex vivo lung perfusions in clinical transplantation. *J Thorac Cardiovasc Surg* 2012;144:1200-6.
90. Lindstedt S, Hlebowicz J, Koul B, et al. Comparative outcome of double lung transplantation using conventional donor lungs and non-acceptable donor lungs reconditioned ex vivo. *Interact Cardiovasc Thorac Surg* 2011;12:162-5.
91. Zych B, Popov AF, Stavri G, et al. Early outcomes of bilateral sequential single lung transplantation after ex-vivo lung evaluation and reconditioning. *J Heart Lung Transplant* 2012;31:274-81.
92. Aigner C, Slama A, Hötzenecker K, et al. Clinical ex vivo lung perfusion—pushing the limits. *Am J Transplant* 2012;12:1839-47.
93. Wallinder A, Ricksten SE, Hansson C, et al. Transplantation of initially rejected donor lungs after ex vivo lung perfusion. *J Thorac Cardiovasc Surg* 2012;144:1222-8.
94. Wallinder A, Ricksten S-E, Silverborn M, et al. Early results in transplantation of initially rejected donor lungs after ex vivo lung perfusion: a case-control study. *Eur J Cardiothorac Surg* 2014;45:40-4.
95. Dark JH, Karamanou D, Clark S, et al. Successful transplantation of unusable donor lungs using ex-vivo lung perfusion: the Newcastle experience (abstract 323). *J Heart Lung Transplant* 2012;31(suppl):S115.
96. Valenza F, Rosso L, Coppola S, et al. Ex vivo lung perfusion to improve donor lung function and increase the number of organs available for transplantation. *Transpl Int* 2014;27:553-61.
97. Henriksen IS, Møller-Sørensen H, Møller CH, et al. First Danish experience with ex vivo lung perfusion of donor lungs before transplantation. *Dan Med J* 2014;61:A4809.
98. Sage E, Mussot S, Trebbia G, et al. Lung transplantation from initially rejected donors after ex vivo lung reconditioning: the French experience. *Eur J Cardiothorac Surg* 2014;46:794-9.
99. Boffini M, Ricci D, Bonato R, et al. Incidence and severity of primary graft dysfunction after lung transplantation using rejected grafts reconditioned with ex vivo lung perfusion. *Eur J Cardiothorac Surg* 2014;46:789-93.
100. Fildes JE, Archer LD, Blaikley J, et al. clinical outcome of patients transplanted with marginal donor lungs via ex vivo lung perfusion compared to standard lung transplantation. *Transplantation* 2015;99:1078-83.
101. Bozso S, Vasanthan V, Luc JG, et al. Lung transplantation from donors after circulatory death using portable ex vivo lung perfusion. *Can Respir J* 2015;22:47-51.
102. Andreasson AS, Dark JH, Fisher AJ. Ex vivo lung perfusion in clinical lung transplantation—state of the art. *Eur J Cardiothorac Surg* 2014;46:779-88.
103. Cypel M, Keshavjee S. Extending the donor pool: rehabilitation of poor organs. *Thorac Surg Clin* 2015;25:27-33.
104. Novel Lung Trial: Normothermic ex vivo lung perfusion (EVLV) as an assessment of extended/marginal donor lungs. <http://clinicaltrials>.

- gov/ct2/show/NCT01365429?term=NOVEL&rank=406/. Accessed April 1, 2016.
105. A study of donor ex-vivo lung perfusion in United Kingdom lung transplantation (DEVELOP-UK). <http://www.isrctn.com/ISRCTN44922411/>. Accessed April 1, 2016.
 106. Fisher A, Andreasson A, Chrysoy A, et al. An observational study of Donor Ex Vivo Lung Perfusion in UK lung transplantation: DEVELOP-UK. *Health Technol Assess* 2016;20:1-276.
 107. International EXPAND Lung Pivotal Trial (EXPAND Lung). <http://clinicaltrials.gov/ct2/show/NCT01963780?term=Expand&rank=42/>. Accessed April 1, 2016.
 108. Extending preservation and assessment time of donor lungs using the Toronto EVLP System™ at a dedicated EVLP facility. <https://clinicaltrials.gov/ct2/show/NCT02234128/>. Accessed April 1, 2016.
 109. Keshavjee SH, Yamazaki F, Cardoso PF, et al. A method for safe twelve-hour pulmonary preservation. *J Thorac Cardiovasc Surg* 1989;98:529-34.
 110. Date H, Matsumura A, Manchester JK, et al. Changes in alveolar oxygen and carbon dioxide concentration and oxygen consumption during lung preservation. *J Thorac Cardiovasc Surg* 1993;105:492-501.
 111. Marczin N, Royston D, Yacoub M. Pro: Lung transplantation should be routinely performed with cardiopulmonary bypass. *J Cardiothorac Vasc Anesth* 2000;14:739-45.
 112. McRae K. Con: lung transplantation should not be routinely performed with cardiopulmonary bypass. *J Cardiothorac Vasc Anesth* 2000;14:746-50.
 113. Aigner C, Wissner W, Taghavi S, et al. Institutional experience with extracorporeal membrane oxygenation in lung transplantation. *Eur J Cardiothorac Surg* 2007;31:468-74.
 114. Ius F, Kuehn C, Tudorache I, et al. Lung transplantation on cardiopulmonary support: venoarterial membrane oxygenation outperformed cardiopulmonary bypass. *J Thorac Cardiovasc Surg* 2012;144:1510-6.
 115. Bermudez CA, Shiose E, Esper SA, et al. Outcomes of intraoperative venoarterial extracorporeal membrane oxygenation versus cardiopulmonary bypass during lung transplantation. *Ann Thorac Surg* 2014;98:1936-42.
 116. Biscotti M, Yang J, Sonett J, et al. Comparison of extracorporeal membrane oxygenation versus cardiopulmonary bypass for lung transplantation. *J Thorac Cardiovasc Surg* 2014;148:2410-5.
 117. Machuca TN, Collaud S, Mercier O, et al. Outcomes of intraoperative extracorporeal membrane oxygenation versus cardiopulmonary bypass for lung transplantation. *J Thorac Cardiovasc Surg* 2015;149:1152-7.
 118. Ius F, Sommer W, Tudorach I, et al. Five-year experience with intraoperative extracorporeal membrane oxygenation in lung transplantation: indications and midterm results. *J Heart Lung Transplant* 2016;35:49-58.
 119. Bhabra MS, Hopkinson DN, Shaw TE, et al. Critical importance of the first 10 minutes of lung graft reperfusion after hypothermic storage. *Ann Thorac Surg* 1996;61:1631-5.
 120. Pierre AF, DeCampos KN, Liu M, et al. Rapid reperfusion causes stress failure in ischemic rat lungs. *J Thorac Cardiovasc Surg* 1998;116:932-42.
 121. Lick SD, Brown PS Jr, Kurusz M, et al. Technique of controlled reperfusion of the transplanted lung in humans. *Ann Thorac Surg* 2000;69:910-2.
 122. Clark SC, Sudarshan CD, Dark JH. Controlled perfusion of the transplanted lung. *Ann Thorac Surg* 2001;71:1755-6.
 123. Schnickel GT, Ross DJ, Beygui R, et al. Modified reperfusion in clinical lung transplantation: the results of 100 consecutive cases. *J Thorac Cardiovasc Surg* 2006;131:218-23.
 124. DeCampos KN, Keshavjee S, Slutsky AS, et al. Alveolar recruitment prevents rapid-reperfusion-induced injury of lung transplants. *J Heart Lung Transplant* 1999;18:1096-102.
 125. Meyers BF, Sundt TM 3rd, Henry S, et al. Selective use of extracorporeal membrane oxygenation is warranted after lung transplantation. *J Thorac Cardiovasc Surg* 2000;120:20-6.
 126. Wigfield CH, Lindsey JD, Steffens TG, et al. Early institution of extracorporeal membrane oxygenation for primary graft dysfunction after lung transplantation improves outcome. *J Heart Lung Transplant* 2007;26:331-8.
 127. Pereszlenyi A, Lang G, Steltzer H, et al. Bilateral lung transplantation with intra- and postoperatively prolonged ECMO support in patients with pulmonary hypertension. *Eur J Cardiothorac Surg* 2002;21:858-63.
 128. Tudorache I, Sommer W, Kühn C, et al. Lung transplantation for severe pulmonary hypertension—awake extracorporeal membrane oxygenation for postoperative left ventricular remodeling. *Transplantation* 2015;99:451-8.
 129. The ARDS Definition Task Force, Ranieri VM, Rubenfeld GD, et al. Acute respiratory distress syndrome: the Berlin definition. *JAMA* 2012;307:2526-33.
 130. Suzuki Y, Cantu E, Christie JD. Primary graft dysfunction. *Semin Respir Crit Care Med* 2013;34:305-19.
 131. Chakinala MM, Kollef MH, Trulock EP. Critical care aspects of lung transplant patients. *J Intens Care Med* 2002;17:8-33.
 132. Fischer S, Bohn D, Rycus P, et al. Extracorporeal membrane oxygenation for primary graft dysfunction after lung transplantation: analysis of the Extracorporeal Life Support Organization (ELSO) registry. *J Heart Lung Transplant* 2007;26:472-7.
 133. Bermudez CA, Adusumilli PS, McCurry KR, et al. Extracorporeal membrane oxygenation for primary graft dysfunction after lung transplantation: long-term survival. *Ann Thorac Surg* 2009;87:854-60.
 134. Hartwig MG, Walczak R, Lin SS, et al. Improved survival but marginal allograft function in patients treated with extracorporeal membrane oxygenation after lung transplantation. *Ann Thorac Surg* 2012;93:366-71.
 135. Beer A, Reed RM, Bölükbas S, et al. Mechanical ventilation after lung transplantation. An international survey of practices and preferences. *Ann Am Thorac Soc* 2014;11:546-53.
 136. Pinsky DJ, Naka Y, Chowdhury NC, et al. The nitric oxide/cyclic GMP pathway in organ transplantation: critical role in successful lung preservation. *Proc Natl Acad Sci USA* 1994;91:23086-90.
 137. Bacha EA, Herve P, Murakami S, et al. Lasting beneficial effect of short term inhaled nitric oxide on graft function after lung transplantation. *J Thorac Cardiovasc Surg* 1996;112:590-8.
 138. Bacha EA, Sellak H, Murakami S, et al. Inhaled nitric oxide attenuates reperfusion injury in non-heart beating-donor lung transplantation. *Transplantation* 1997;63:1380-6.
 139. Bhabra MS, Hopkinson DN, Shaw TE, et al. Low dose nitric oxide inhalation during initial reperfusion enhances rat lung graft function. *Ann Thorac Surg* 1997;63:339-44.
 140. Strüber M, Harringer W, Ernest M, et al. Inhaled nitric oxide as a prophylactic treatment against reperfusion injury of the lung. *Thorac Cardiovasc Surg* 1999;47:179-82.
 141. Meade MO, Granton JT, Matte-Martyn A, et al. A randomized trial of inhaled nitric oxide to prevent ischemia-reperfusion injury after lung transplantation. *Am J Respir Crit Care Med* 2003;167:1483-9.
 142. Perrin G, Roch A, Michelet P, et al. Inhaled nitric oxide does not prevent pulmonary edema after lung transplantation measured by lung water content: a randomized clinical study. *Chest* 2006;129:1024-30.
 143. Botha P, Jeyakanthan M, Rao JN, et al. Inhaled nitric oxide for modulation of ischemia-reperfusion injury in lung transplantation. *J Heart Lung Transplant* 2007;26:1199-205.
 144. Liu Y, Liu Y, Su L, et al. Recipient-related clinical risk factors for primary graft dysfunction after lung transplantation: a systematic review and meta-analysis. *PLoS One* 2014;9:e92773-84.
 145. Rossaint R, Falke KJ, Lopez F, et al. Inhaled nitric oxide for the adult respiratory distress syndrome. *N Engl J Med* 1993;328:399-405.
 146. Adatia I, Lillehei C, Arnold JH, et al. Inhaled nitric oxide in the treatment of postoperative graft dysfunction after lung transplantation. *Ann Thorac Surg* 1994;57:1311-8.
 147. MacDonald P, Mundy J, Roger P, et al. Successful treatment of life-threatening acute reperfusion injury after lung transplantation with inhaled nitric oxide. *J Thorac Cardiovasc Surg* 1995;110:861-3.
 148. Date H, Triantafyllou AN, Trulock EP, et al. Inhaled nitric oxide reduces human lung allograft dysfunction. *J Thorac Cardiovasc Surg* 1996;111:913-9.
 149. Garat C, Jayr C, Eddahibi S, et al. Effects of inhaled nitric oxide or inhibition of endogenous nitric oxide formation on hypoxic lung injury. *Am J Respir Crit Care Med* 1997;155:1957-64.

150. Fuller BM, Mohr NM, Skrupky L, et al. The use of inhaled prostaglandins in patients with ARDS. *Chest* 2015;147:1510-22.
151. Walmrath D, Schneider T, Schermuly R, et al. Direct comparison of inhaled nitric oxide and aerosolized prostacyclin in acute respiratory distress syndrome. *Am J Respir Crit Care Med* 1996;153:991-6.
152. Fiser SM, Cope JT, Kron IL, et al. Aerosolized prostacyclin (epoprostenol) as an alternative to inhaled nitric oxide for patients with reperfusion injury after lung transplantation. *J Thorac Cardiovasc Surg* 2001;121:981-2.
153. Khan TA, Schnickel G, Ross D, et al. A prospective, randomized, crossover pilot study of inhaled nitric oxide versus inhaled prostacyclin in heart transplant and lung transplant recipients. *J Thorac Cardiovasc Surg* 2009;138:1417-24.
154. Kawashima M, Nakamura T, Schneider S, et al. Iloprost ameliorates post-ischemic lung reperfusion injury and maintains an appropriate pulmonary ET-1 balance. *J Heart Lung Transplant* 2003;22:794-801.
155. Wittwer T, Franke UFW, Fehrenbach A, et al. Donor pretreatment using the aerosolized prostacyclin analogue iloprost optimizes post-ischemic function and non-heart beating donor lungs. *J Heart Lung Transplant* 2005;24:371-8.
156. Hartwig MG, Appel JZ 3rd, Cantu E 3rd, et al. Improved results treating lung allograft failure with venovenous extracorporeal membrane oxygenation. *Ann Thorac Surg* 2005;80:1872-9.
157. Yusen RD, Edwards LB, Kucheryavaya AY, et al. The Registry of the ISHLT: thirty-first adult lung and heart-lung transplant report—2014; Focus theme: Retransplantation. *J Heart Lung Transplant* 2014;33:1009-24.
158. Aigner C. Retransplantation. *Curr Opin Organ Transplant* 2015;20:521-6.
159. Osho AA, Castleberry AW, Snyder LD, et al. Differential outcomes with early and late repeat transplantation in the era of the lung allocation score. *Ann Thorac Surg* 2014;98:1914-20.
160. Griese M. Pulmonary surfactant in health and human lung diseases: state of the art. *Eur Respir J* 1999;13:1455-76.
161. Whitsett JA. The molecular era of surfactant biology. *Neonatology* 2014;105:337-43.
162. Günther A, Balsler M, Schmidt R, et al. Surfactant abnormalities after single lung transplantation in dogs: impact of bronchoscopic surfactant administration. *J Thorac Cardiovasc Surg* 2004;127:344-54.
163. Friedrich I, Börgermann J, Splittgerber FH, et al. Bronchoscopic surfactant administration preserves gas exchange and pulmonary compliance after single lung transplantation in dogs. *J Thorac Cardiovasc Surg* 2004;127:335-43.
164. Erasmus ME, Petersen AH, Oetomo SB, et al. The function of surfactant is impaired during the reimplantation response in rat lung transplants. *J Heart Lung Transplant* 1994;13:791-802.
165. Erasmus ME, Petersen AH, Hofstede G, et al. Surfactant treatment before reperfusion improves the immediate function of lung transplants in rats. *Am J Respir Crit Care Med* 1996;153:665-70.
166. Andrade RS, Solien EE, Wangenstein OD, et al. Surfactant dysfunction in lung preservation. *Transplantation* 1995;60:536-41.
167. Hohlfeld JM, Tiryaki E, Hamm H, et al. Pulmonary surfactant activity is impaired in lung transplant recipients. *Am J Respir Crit Care Med* 1998;158:706-12.
168. Waldhausen JA, Giammona ST, Kilman JW, et al. Effect of transplantation of canine lung on pulmonary compliance and surfactant. *JAMA* 1965;191:1002-5.
169. Ochs M, Fehrenbach H, Nenadic I, et al. Preservation of intraalveolar surfactant in a rat lung ischaemia/reperfusion injury model. *Eur Respir J* 2000;15:526-31.
170. Veldhuizen RA, Lee J, Sandler D, et al. Alterations in pulmonary surfactant composition and activity after experimental lung transplantation. *Am Rev Respir Dis* 1993;148:208-15.
171. Koletsis E, Chatzimichalis A, Fotopoulos V, et al. Donor lung pretreatment with surfactant in experimental transplantation preserves graft hemodynamics and alveolar morphology. *Exp Biol Med* (Maywood) 2003;228:540-5.
172. Novick RJ, Veldhuizen RA, Possmayer F, et al. Exogenous surfactant therapy in thirty-eight hour lung graft preservation for transplantation. *J Thorac Cardiovasc Surg* 1994;108:259-68.
173. Novick RJ, MacDonald J, Veldhuizen RA, et al. Evaluation of surfactant treatment strategies after prolonged graft storage in lung transplantation. *Am J Respir Crit Care Med* 1996;154:98-104.
174. Novick RJ, Gilpin AA, Gehman KE, et al. Mitigation of injury in canine lung grafts by exogenous surfactant therapy. *J Thorac Cardiovasc Surg* 1997;113:342-53.
175. Hausen B, Rohde R, Hewitt CW, et al. Exogenous surfactant treatment before and after sixteen hours of ischemia in experimental lung transplantation. *J Thorac Cardiovasc Surg* 1997;113:1050-8.
176. Hohlfeld JM, Strüber M, Ahlf K, et al. Exogenous surfactant improves survival and surfactant function in ischaemia-reperfusion injury in minipigs. *Eur Respir J* 1999;13:1037-43.
177. Maitra G, Inchley K, Novick RJ, et al. Acute lung injury and lung transplantation influence in vitro subtype conversion of pulmonary surfactant. *Am J Physiol Lung Cell Mol Physiol* 2002;282:L67-74.
178. Erasmus ME, Hofstede GJ, Petersen AH, et al. SP-A-enriched surfactant for treatment of rat lung transplants with SP-A deficiency after storage and reperfusion. *Transplantation* 2002;73:348-52.
179. Strüber M, Cremer J, Harringer W, et al. Nebulized synthetic surfactant in reperfusion injury after single lung transplantation. *J Thorac Cardiovasc Surg* 1995;110:563-4.
180. Strüber M, Hirt SW, Cremer J, et al. Surfactant replacement in reperfusion injury after clinical lung transplantation. *Intensive Care Med* 1999;25:862-4.
181. Warnecke G, Strüber M, Fraud S, et al. Combined exogenous surfactant and inhaled nitric oxide therapy for lung ischaemia-reperfusion injury in minipigs. *Transplantation* 2001;71:1238-44.
182. Strüber M, Hohlfeld JM, Kofidis T, et al. Surfactant function in lung transplantation after 24 hours of ischemia: advantage of retrograde flush perfusion for preservation. *J Thorac Cardiovasc Surg* 2002;123:98-103.
183. Mühlfeld C, Becker L, Bussinger C, et al. Exogenous surfactant in ischemia/reperfusion: effects on endogenous surfactant pools. *J Heart Lung Transplant* 2010;29:327-34.
184. van der Kaaij NP, Haitsma JJ, Kluin J, et al. Surfactant pretreatment ameliorates ischemia-reperfusion injury of the lung. *Eur J Cardiothorac Surg* 2005;27:774-82.
185. van der Kaaij NP, Kluin J, Haitsma JJ, et al. Surfactant pretreatment decreases long-term damage after ischemia-reperfusion injury of the lung. *Eur J Cardiothorac Surg* 2009;35:304-12.
186. Dreyer N, Mühlfeld C, Fehrenbach A, et al. Exogenous surfactant application in a rat lung ischemia reperfusion injury model: effects on edema formation and alveolar type II cells. *Respir Res* 2008;9:5.
187. Wittwer T, Madershahian N, Rahmanian P, et al. Surfactant application in experimental lung transplantation. *J Heart Lung Transplant* 2013;32:355-9.
188. Strüber M, Fischer S, Niedermeyer J, et al. Effects of exogenous surfactant instillation in clinical lung transplantation: a prospective, randomized trial. *J Thorac Cardiovasc Surg* 2007;133:1620-5.
189. Amital A, Shitrit D, Raviv Y, et al. The use of surfactant in lung transplantation. *Transplantation* 2008;86:1549-56.
190. Kermeen FD, McNeil KD, Fraser JF, et al. Resolution of severe ischemia-reperfusion injury post-lung transplantation after administration of endobronchial surfactant. *J Heart Lung Transplant* 2007;26:850-6.
191. Amital A, Shitrit D, Raviv Y, et al. Surfactant as salvage therapy in life threatening primary graft dysfunction in lung transplantation. *Eur J Cardiothorac Surg* 2009;35:299-303.
192. Scherer M, Demertzis S, Langer F, et al. C1-esterase inhibitor reduces reperfusion injury after lung transplantation. *Ann Thorac Surg* 2002;73:233-8.
193. Keshavjee S, Davis RD, Zamora MR, et al. A randomized, placebo-controlled trial of complement inhibition in ischemia-reperfusion injury after lung transplantation in human beings. *J Thorac Cardiovasc Surg* 2005;129:423-8.
194. Pierre AF, Xavier AM, Liu M, et al. Effect of complement inhibition with soluble complement receptor 1 on pig allotransplant lung function. *Transplantation* 1998;66:723-32.

195. Schmid RA, Zollinger A, Singer T, et al. Effect of soluble complement receptor type 1 on reperfusion edema and neutrophil migration after lung allotransplantation in swine. *J Thorac Cardiovasc Surg* 1998;116:90-7.
196. Strüber M, Hagl C, Hirt SW, et al. CI-esterase inhibitor in graft failure after lung transplantation. *Intens Care Med* 1999;25:1315-8.
197. Zamora MR, Davis RD, Keshavjee SH, et al. Complement inhibition attenuates human lung transplant reperfusion injury: a multicenter trial. *Chest* 1999;116(suppl):46S.
198. Stammberger U, Hamacher J, Hillinger S, et al. sCR1sLe ameliorates ischemia/reperfusion injury in experimental lung transplantation. *J Thorac Cardiovasc Surg* 2000;120:1078-84.
199. Stammberger U, Hamacher J, Pache JC, et al. sCR1sLe(X) reduces lung allograft ischemia-reperfusion injury but does not ameliorate acute rejection. *Eur J Cardiothorac Surg* 2002;22:368-72.
200. Khan MA, Nicolls MR. Complement-mediated microvascular injury leads to chronic rejection. *Adv Exp Med Biol* 2013;735:233-46.
201. Damman J, Hoeger S, Boneschansker L, et al. Targeting complement activation in brain-dead donors improves renal function after transplantation. *Transpl Immunol* 2011;24:233-7.
202. Conte JV Jr, Katz NM, Wallace RB, et al. Long-term lung preservation with the PAF antagonist BN 52021. *Transplantation* 1991;51:1152-6.
203. Qayumi AK, Jamieson WR, Poostizadeh A. Effects of platelet-activating factor antagonist CV-3988 in preservation of heart and lung for transplantation. *Ann Thorac Surg* 1991;52:1026-32.
204. Hirt SW, Wahlers T, Jurmann M, et al. Antagonisation of platelet activating factor—a new therapeutic concept for improvement of organ quality in lung preservation. *Transpl Int* 1992;5(suppl 1):S374-8.
205. Wahlers T, Hirt SW, Haverich A, et al. Future horizons of lung preservation by application of a platelet-activating factor antagonist compared with current clinical standards. Euro-Collins flush perfusion versus donor core cooling. *J Thorac Cardiovasc Surg* 1992;103:200-4.
206. Corcoran PC, Wang Y, Katz NM, et al. Platelet activating factor antagonist enhances lung preservation. *J Surg Res* 1992;52:615-20.
207. Corcoran PC, Wang Y, Katz NM, et al. Platelet activating factor antagonist enhances lung preservation in a canine model of single lung allotransplantation. *J Thorac Cardiovasc Surg* 1992;104:66-72.
208. Kawahara K, Tagawa T, Takahashi T, et al. The effect of the platelet-activating factor inhibitor TCV-309 on reperfusion injury in a canine model of ischemic lung. *Transplantation* 1993;55:1438-9.
209. Stammberger U, Carboni GL, Hillinger S, et al. Combined treatment with endothelin- and PAF-antagonists reduces posttransplant lung ischemia/reperfusion injury. *J Heart Lung Transplant* 1999;18:862-8.
210. Wittwer T, Grote M, Oppelt P, et al. Impact of PAF antagonist BN 52021 (Ginkgolide B) on post-ischemic graft function in clinical lung transplantation. *J Heart Lung Transplant* 2001;20:358-63.
211. Grino JM. BN 52021: a platelet activating factor antagonist for preventing post-transplant renal failure. A double-blind, randomized study. The BN 52021 Study Group in Renal Transplantation. *Ann Intern Med* 1994;121:345-7.
212. Beuk RJ, Tangelder GJ, Maassen RL, et al. Leucocyte and platelet adhesion in different layers of the small bowel during experimental total warm ischaemia and reperfusion. *Br J Surg* 2008;95:1294-304.
213. Zarbock A, Singbartl K, Ley K. Complete reversal of acid-induced acute lung injury by blocking of platelet-neutrophil aggregation. *J Clin Invest* 2006;116:3211-9.
214. Looney MR, Nguyen JX, Hu Y, et al. Platelet depletion and aspirin treatment protect mice in a two-event model of transfusion-related acute lung injury. *J Clin Invest* 2009;119:3450-61.
215. Boyle AJ, Di Gangi S, Hamid UI, et al. Aspirin therapy in patients with acute respiratory distress syndrome (ARDS) is associated with reduced intensive care unit mortality: a prospective analysis. *Crit Care* 2015;19:109.
216. Akinosoglou K, Alexopoulos D. Use of antiplatelet agents in sepsis: a glimpse into the future. *Thromb Res* 2014;133:131-8.
217. Sayah DM, Mallavia B, Liu F, et al. Neutrophil extracellular traps are pathogenic in primary graft dysfunction after lung transplantation. *Am J Respir Crit Care Med* 2015;191:455-63.
218. Bosmann M, Ward PA. Protein-based therapies for acute lung injury: targeting neutrophil extracellular traps. *Expert Opin Ther Targets* 2014;18:703-14.
219. La Francesca S, Ting AE, Sakamoto J, et al. Multipotent adult progenitor cells decrease cold ischemic injury in ex vivo perfused human lungs: an initial pilot and feasibility study. *Transplant Res* 2014;3:19.
220. Bustos ML, Huleihel L, Meyer EM, et al. Activation of human mesenchymal stem cells impacts their therapeutic abilities in lung injury by increasing interleukin (IL)-10 and IL-1RN levels. *Stem Cells Transl Med* 2013;2:884-95.
221. Simonson OE, Mouggiakakos D, Heldring N, et al. In vivo effects of mesenchymal stromal cells in two patients with severe acute respiratory distress syndrome. *Stem Cells Transl Med* 2015;4:1199-213.
222. Liu KD, Wilson JG, Zhuo H, et al. Design and implementation of the START (STem cells for ARDS Treatment) trial, a phase 1/2 trial of human mesenchymal stem/stromal cells for the treatment of moderate-severe acute respiratory distress syndrome. *Ann Intensive Care* 2014;4:22.
223. Tiriveedhi V, Gautam B, Sarma NJ, et al. Pre-transplant antibodies to α 1 tubulin and collagen-V in lung transplantation: clinical correlations. *J Heart Lung Transplant* 2013;32:807-14.
224. Cantu E, Shah RJ, Lin W, et al. Oxidant stress regulatory genetic variation in recipients and donors contributes to risk of primary graft dysfunction after lung transplantation. *J Heart Lung Transplant* 2014;33:1093-9.
225. Somers J, Rutten D, Verleden SE, et al. Interleukin-17 receptor polymorphism predisposes to primary graft dysfunction after lung transplantation. *J Heart Lung Transplant* 2015;34:941-9.
226. Perkins GD, Gates S, Park D, et al. The beta agonist lung injury trial prevention. A randomised, controlled trial. *Am J Respir Crit Care Med* 2014;189:674-83.
227. McAuley DF, Laffey JG, O'Kane CM, et al. Simvastatin in the acute respiratory distress syndrome. *N Engl J Med* 2014;371:1695-703.
228. Yusen RD, Edwards LB, Kucheryavaya AY, et al. The Registry of the International Society for Heart and Lung Transplantation: thirty-second adult lung and heart-lung transplant report—2015; Focus theme: Early graft failure. *J Heart Lung Transplant* 2015;34:1264-77.
229. Lucas R, Verin AD, Black SM, et al. Regulators of endothelial and epithelial barrier integrity and function in acute lung injury. *Biochem Pharmacol* 2009;77:1763-72.